Detection (FISH)

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Introduction

Probes labeled with biotin must be detected with fluorescently labeled Avidin, and probes labeled with digoxigenin require detection with a fluorochrome conjugated antibody against this hapten. For example, to detect the biotin labeled probes we routinely use Avidin-FITC, Avidin-TRITC, or Avidin Cy-5. For the probes labeled with digoxigenin, we usually first incubate with mouse-anti-digoxigenin, followed by incubation with sheep anti-mouse Cy5.5, or other fluorochrome conjugated antibodies.

Reagents

Avidin-Cy5

Jackson Immuno Research Lab, Cat. 003-170-083

Avidin-TRITC

Sigma, Cat. A 7169

Avidin-FITC

Vector, Cat. A-2011

BSA (Bovine Serum Albumin)

DAPI

Ethanol, absolute

Formamide

Fluka BioChemika, Cat. 47671

HCl, 1N

Mouse anti-digoxigenin

Sigma, Cat. D 8156

Sheep anti-mouse Cy5.5

Amersham, Cat. RPQ 0115

20X SSC

Tween 20

Preparation of Reagents

50% FA/SSC

20X SSC 30 ml dH_2O 120 ml Formamide 150 ml Adjust pH to 7 with 1N HCl

Pre-warm to 45°C

1X SSC (for direct labeled probes, i.e., TRITC, FITC or other)

 $\begin{array}{ccc} 20X \ SSC & 25 \ ml \\ dH_2O & 475 \ ml \end{array}$

Pre-warm to 45°C

0.1X SSC (for indirect labeled probes, i.e. Biotin, or Digoxigenin)

20X SSC 2.5 ml dH2O 497.5 ml

Pre-warm to 60°C

4X SSC/0.1%Tween20

 $\begin{array}{ccc} 20 X \ SSC & 200 \ ml \\ dH_2O & 799 \ ml \\ Tween \ 20 & 1 \ ml \end{array}$

Pre-warm to 45°C

Blocking Solution (3% BSA/4X SSC/0.1%Tween20)

BSA 0.3 g 4X SSC/0.1%Tween 20 10 ml

Pre-warm to 37°C

Antibody Solution (1% BSA/4X SSC/0.1%Tween 20)

BSA 0.1 g 4X SSC/0.1%Tween 20 10 ml

Pre-warm to 37°C

DAPI stock solution (f.c.= 0.2 mg/ml)

 $\begin{array}{cc} \mathrm{DAPI} & 2 \ \mathrm{mg} \\ \mathrm{ddH_2O} & 10 \ \mathrm{ml} \end{array}$

Aliquot and store at -80°C

DAPI staining solution (f.c.= 80 ng/ml)

DAPI (stock solution) 40 µl 2X SSC 100 ml Store at 4°C in a light-tight coplin jar

Procedure

- 1. Carefully remove the rubber cement surrounding the coverslips from hybridized slides.
- 2. Wash the slides in 50% formamide/2X SSC (pH 7-7.5) for 3 x 5 min at 45°C, shaking.
- 3. Wash slides in 0.1X SSC at 60°C (for indirectly labeled probes) or 1X SSC at 45°C (for directly labeled probes) for 3 x 5 min, shaking.
- 4. Dip slides in 4X SSC/0.1%Tween 20.
- 5. Add 120 μl of Blocking Solution (3% BSA/4X SSC/0.1%Tween 20) to the slides and cover them with a 24 mm x 60 mm coverslip in a moist hybridization chamber at 37°C for 30 min.
- 6. Dip slides in 4X SSC/0.15Tween 20 to wash off the blocking solution. Proceed directly to step 9 if using a directly-labeled probe.
- 7. For indirectly-labeled probes (Biotin or Digoxigenin), add 120 μl of fluorescent antibody (antibody should be diluted 1:200 in 1% BSA/4X SSC/0.1%Tween 20) to the slides, cover with a 24 mm x 60 mm coverslip, and incubate in moist light-tight hybridization chamber at 37°C for 45 min.
- 8. Wash slides in 4X SSC/0.1%Tween 20, for 3 x 5 min, shaking.
- 9. Stain slides for 5 min in DAPI staining solution in a light-protected coplin jar.
- 10. Wash the slides for 5 min in 2X SSC, shaking.
- Dehydrate the slides by dipping through an ethanol series of: 70%, 90%, and 100%; air-dry.
- 12. Apply 35 μl of antifade solution, cover with 24 mm x 60 mm coverslips, store in light-protected container at 4°C until slide is imaged.

Notes

1. Exposure of slides to ambient light should be minimized during all procedures.

- 2. Use care in removing coverslips during all procedures to minimize scratches.
- 3. Spin all fluorescent dyes prior to use for 3 min at 13,000 rpm and carefully pipette the antibody without disturbing the pellet.
- 4. Do not let the slide dry out between washing steps.